



# TAL TECH

## SENSORS LECTURE 8

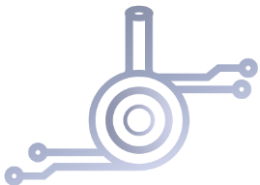
Tamás Pardy  
TalTech Lab-on-a-Chip

01.02.2022

**TALLINN UNIVERSITY  
OF TECHNOLOGY**

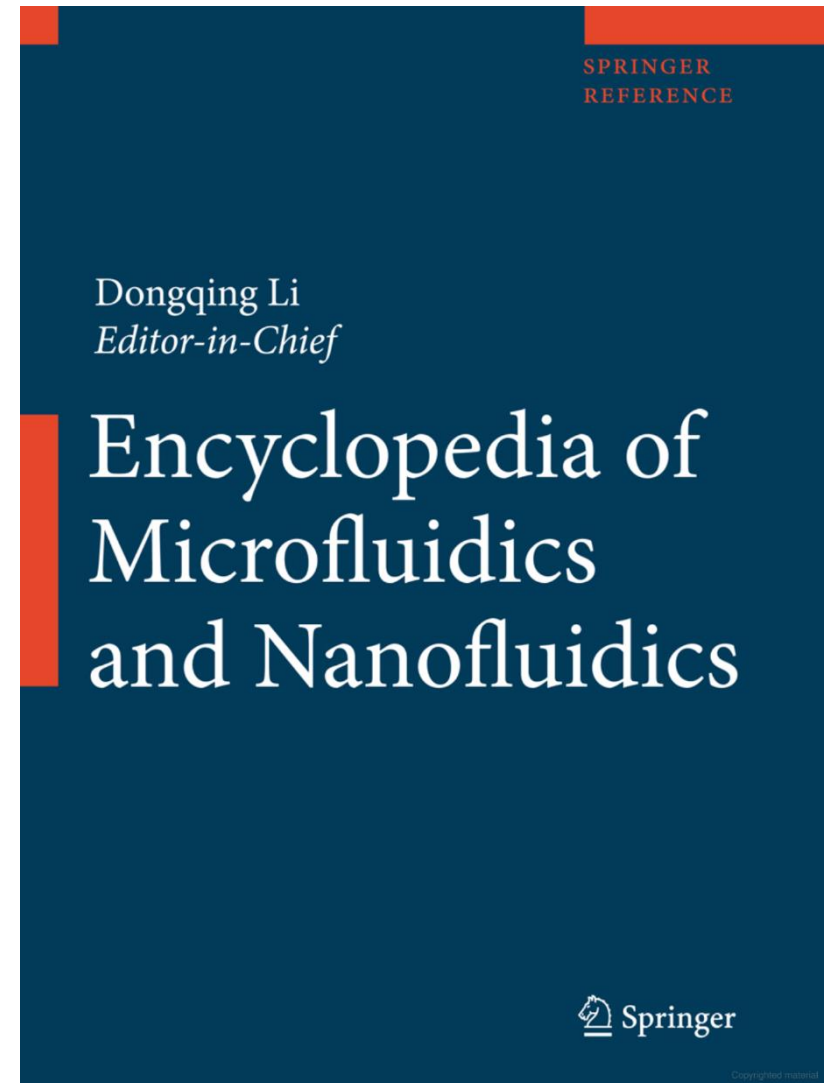
## BEFORE WE BEGIN...

- Encyclopedia of Microfluidics and Nanofluidics
- Dongqing Li
- ISBN: 9780387324685
- Slides based on originals from IEE1720 lecture 1 by Natalja Sleptšuk
- BioMEMS course, Peter Pazmany Catholic University, Budapest, Hungary (by: Kristof Ivan)
- **Additional materials:**
  - <https://www.sciencedirect.com/science/article/pii/S0956566315303298>



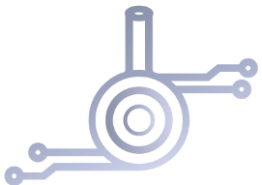
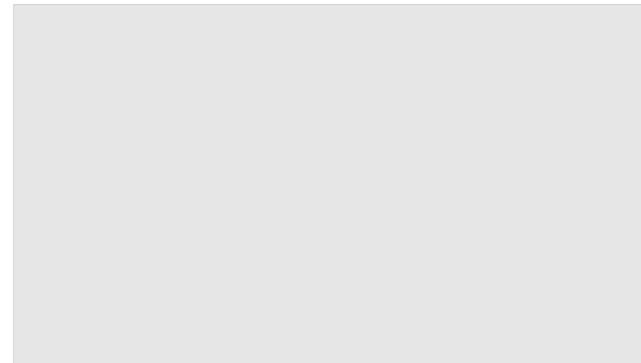
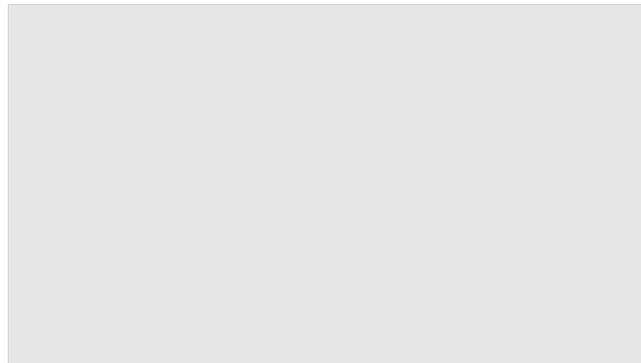
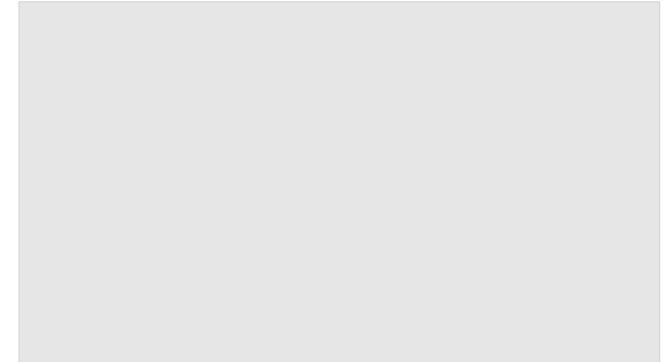
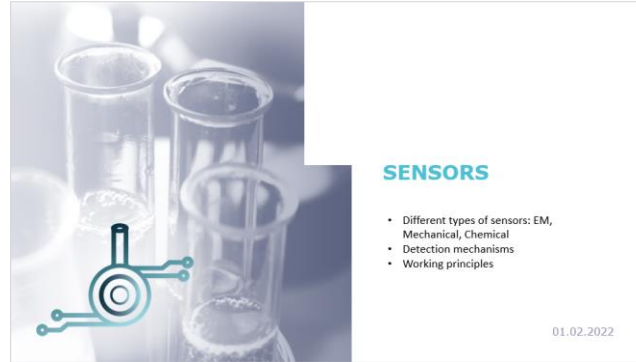
**TAL  
TECH**

TALLINN UNIVERSITY  
OF TECHNOLOGY



01.02.2022

# OVERVIEW





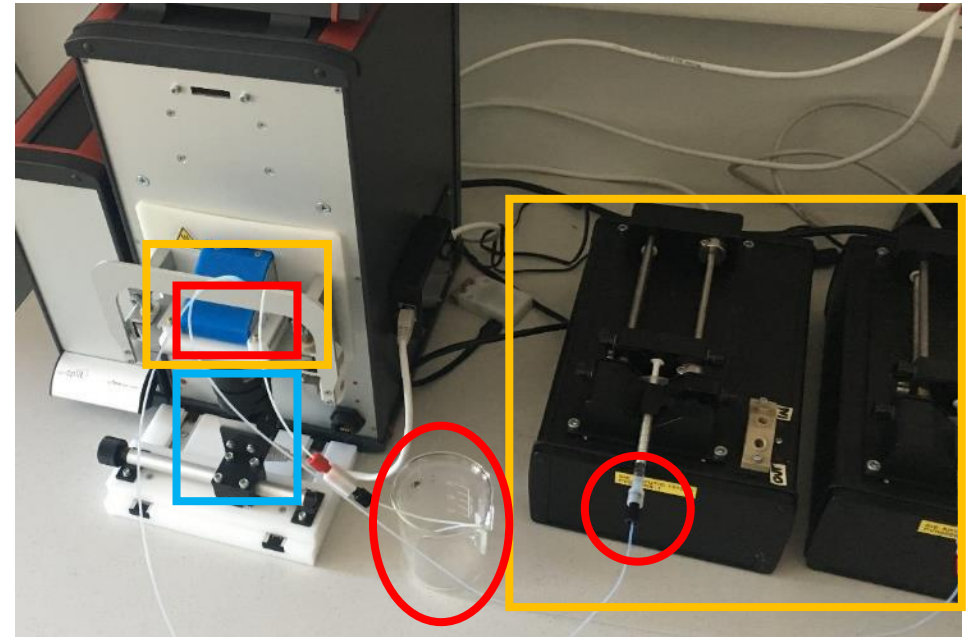
# INTRODUCTION

- What are sensors?
- How do biosensors work?
- Human biomarkers

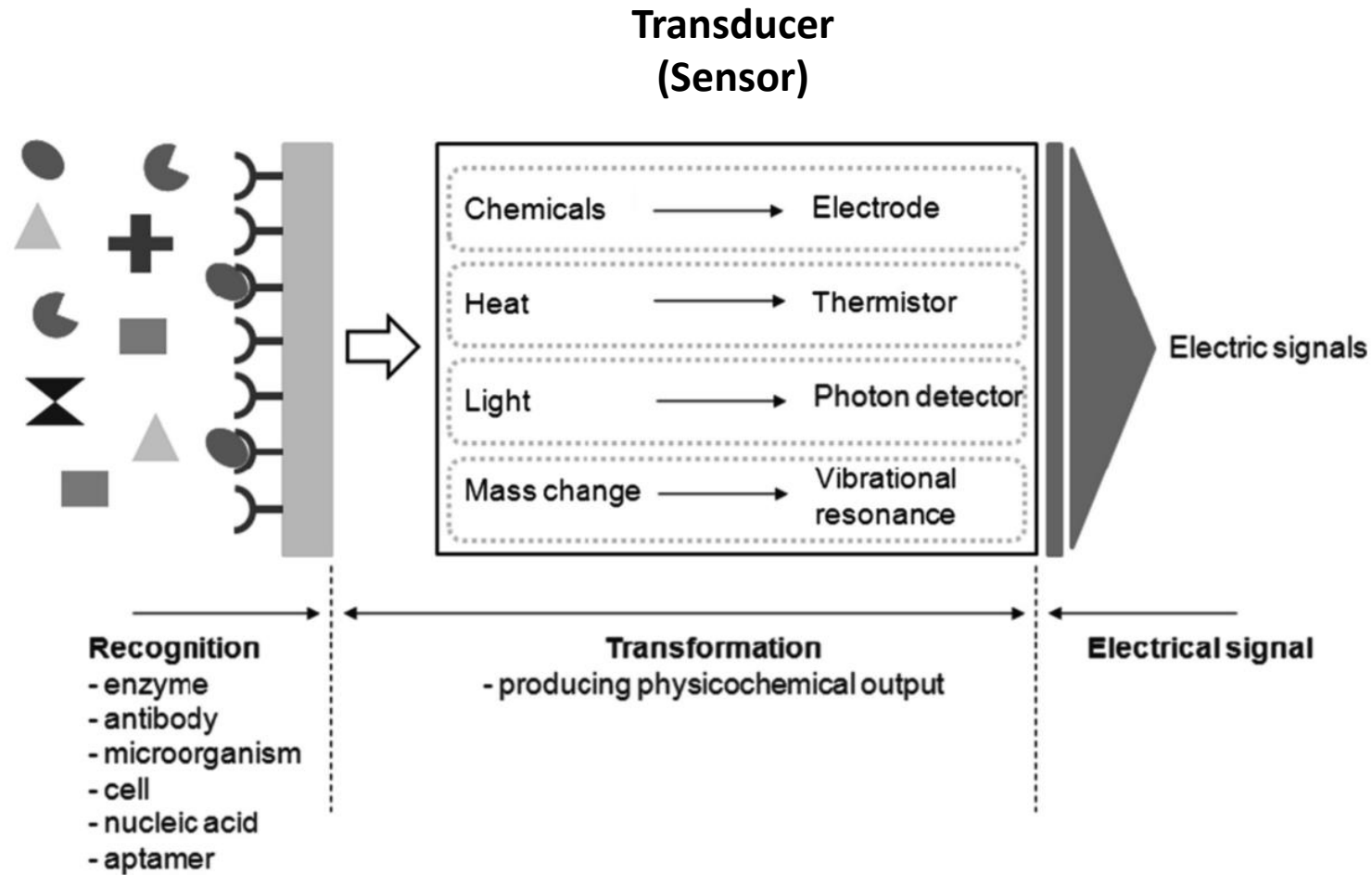
01.02.2022

# EXTERNAL SYSTEM COMPONENTS

- 1. **Liquid path:** → Lecture 3
  - Liquid reservoir
  - Tubing, fittings, chip holder, connector
  - Microfluidic chip (reactor)**
  - Product/waste collector
- 2. **Actuators (internal/external):** → Lecture 9
  - Pump, valve, flow regulator
  - Thermal regulation (thermostat, heater etc.)
- 3. **Sensors (internal/external):**
  - Optical: camera, fluorometer etc.
  - Electrical: impedimetric sensors
  - Flow control: pressure sensor, flow meter
- Plus, power supply, control board, network interface, user I/O interfaces etc.



# SENSORS



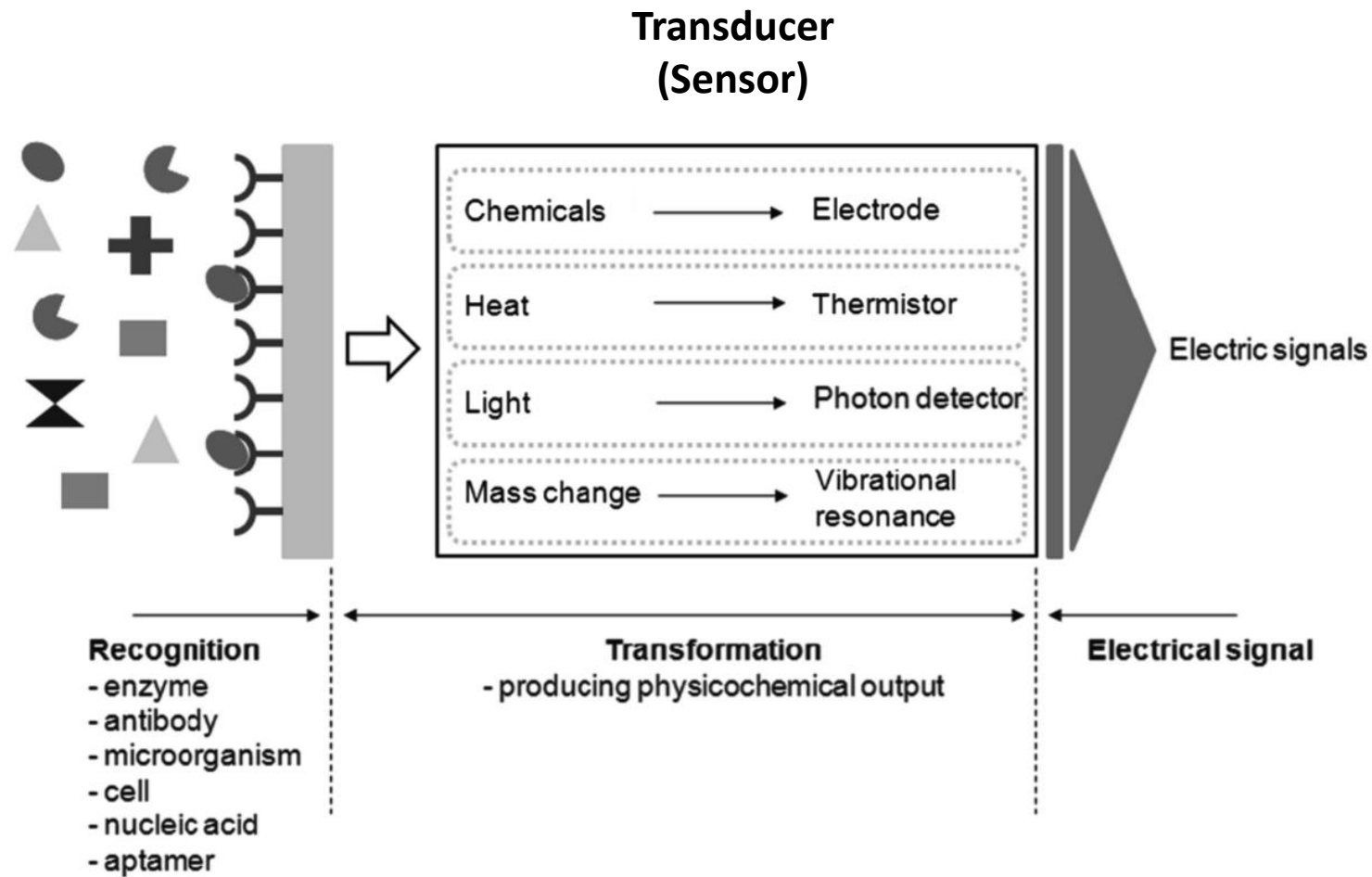
Sensor:

“**converts** one form of **energy** to another, and in so doing detects and conveys information about some physical, chemical or biological phenomena”

Sensor, as **transducer**:

converts the **measurand** (a quantity or a parameter) into a signal that carries information

# SENSORS



## Ideal sensor

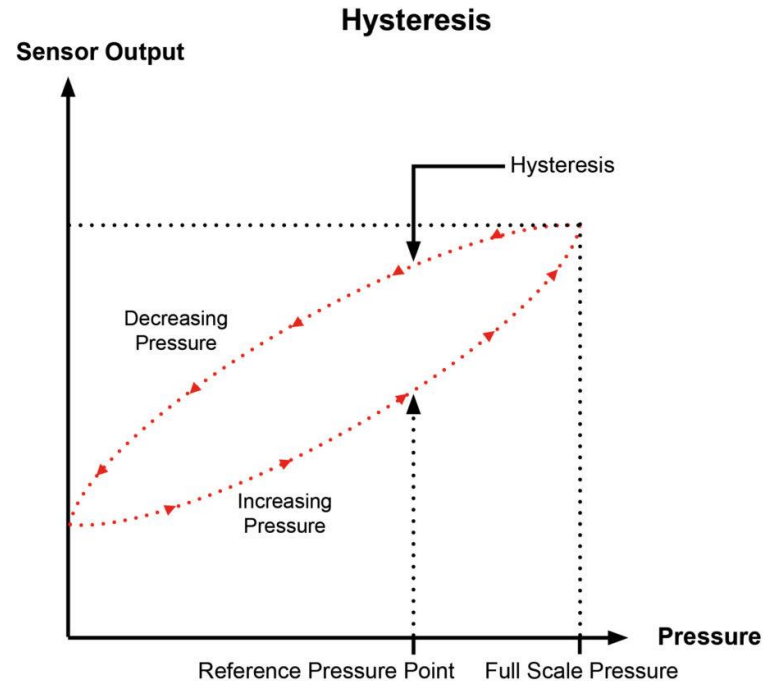
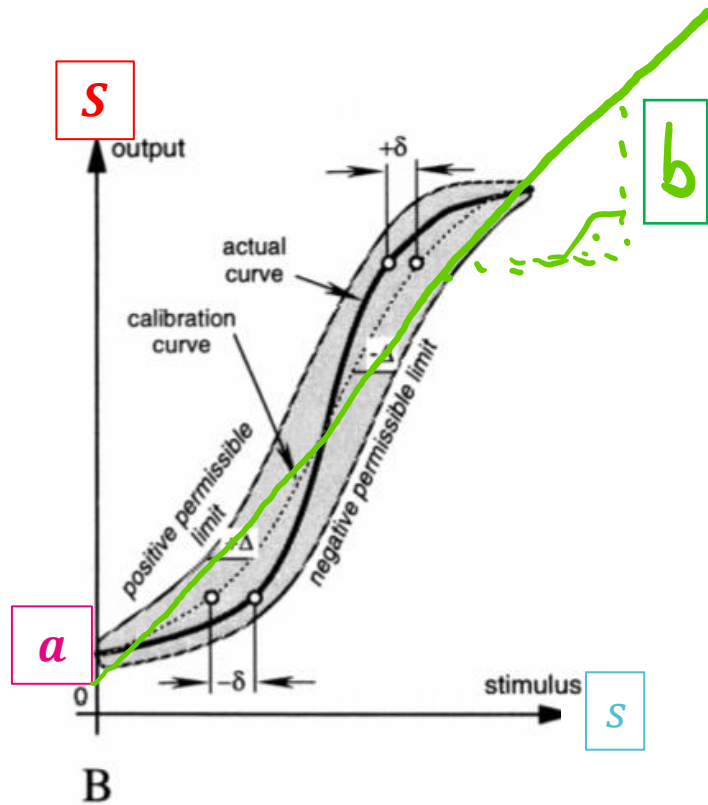
- Measures continuously
- Is sensitive and selective
- Has fast and predictable response
- Has reversible behavior
- High SNR
- Compact
- Immune to environmental interference
- Is easy to calibrate

## Categories

- Biosensor
- Smart sensor
- Passive/active
- Array type
- Multimodal...



# SENSORS



## Transfer function:

(Relation between output signal and measurand)

$$S = a + bs$$

S: output electric signal

a: intercept (output at zero input signal)

b: slope/sensitivity

s: input stimulus

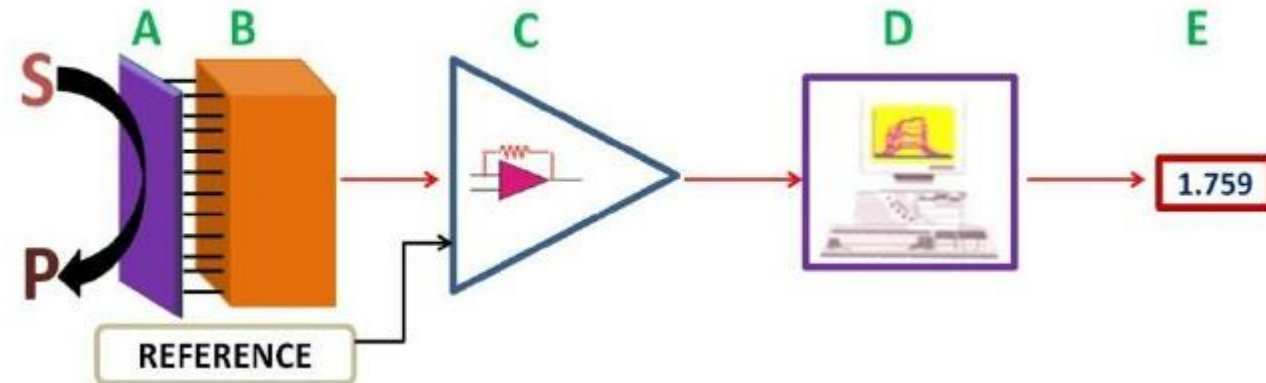
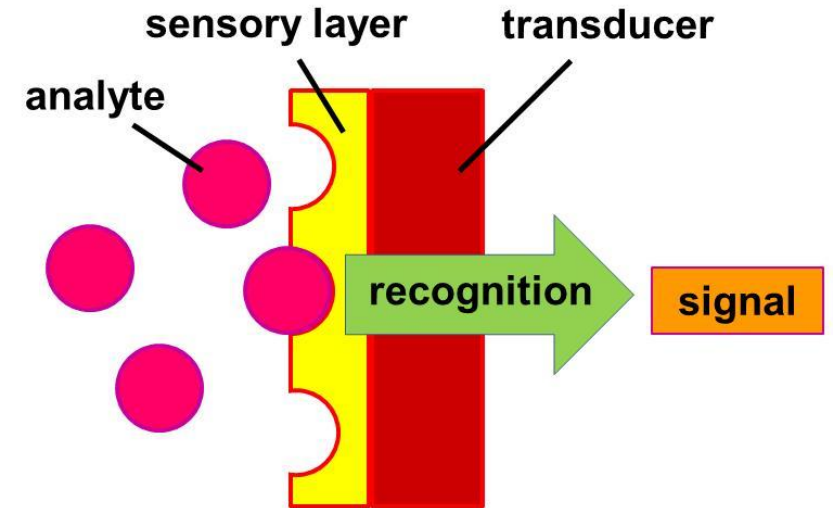
## Hysteresis (error):

Maximum difference between outputs at a specific point of input stimulus when approached first increasing then decreasing stimulus (e.g.: inflating and deflating a balloon while measuring pressure)



# WHAT IS A BIOSENSOR?

- **Goal:** to tell how much of something you have in a solution
- Something = target/analyte, measured in **concentration** (e.g. molarity in  $\text{mol}/\text{m}^3$  or  $\text{mol}/\text{L}$ )
- **Principle**
  - A: Reaction, produces product P in a specific concentration
  - B: transducer converts concentration to electric signal
  - C: signal is amplified
  - D: signal is processed
  - E: processed, amplified signal is visualized in human-readable form

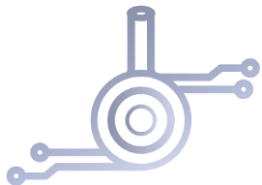


# HUMAN BIOMARKERS

- **Biomarker** = "chemical, its metabolite, or the product of an interaction between a chemical and some target molecule or cell that is measured in the human body" (WHO definition)

Biomarker concentration  $\Leftrightarrow$  physiological state of organism

- **Example:**
  - hCG as indicator of pregnancy
  - Released by placenta after implantation of fertilized egg
  - Rapid tests available since 60's, today <1€/test



Category	Detection target	Bodily fluid	Biomarker or substance
Cancer detection	prostate cancer	blood	PSA (prostate specific antigen)
	liver cancer	blood	AFP (tumor marker alpha-fetoprotein)
Other commonly used	pregnancy	urine	hCG (human chorionic gonadotropin)
Cardiac markers	myocardial damage (infarction)	blood	troponin
	myocardial infarction (no skeletal muscle damage)	blood	creatine kinase (CK-MB)
	myocardial infarction	blood	lactate dehydrogenase (LDH-1)
Exposure to toxic substance (biomonitoring)	lead	blood	blood lead
	arsenic	urine	urinary arsenic
	nitrites	blood	methemoglobin
	phthalates	urine	phthalate metabolites

Source: (own work) course work report, IEM9040, by Tamas Pardy, 2014



# INTRODUCTION

- What are sensors?
- How do biosensors work?
- Human biomarkers

01.02.2022



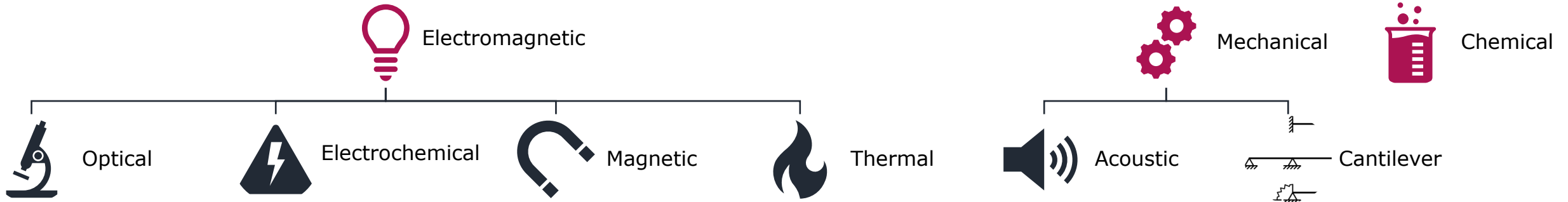
# SENSORS

- Different types of sensors: EM, Mechanical, Chemical
- Detection mechanisms
- Working principles

01.02.2022

# DETECTION METHODS

## Technology



### Labeling

- Labeled
- Label-free

### Amplification

- Signal amplification
- Target amplification

### State of organism

- In vivo
- In vitro
- *Ex vivo (e.g. cell cultures)*

# SPECTROPHOTOMETRY

- Beer-Lambert Law:  
attenuation of light  $\Leftrightarrow$  properties of material being lit

$$A = \epsilon l c \text{ [a.u.]}$$

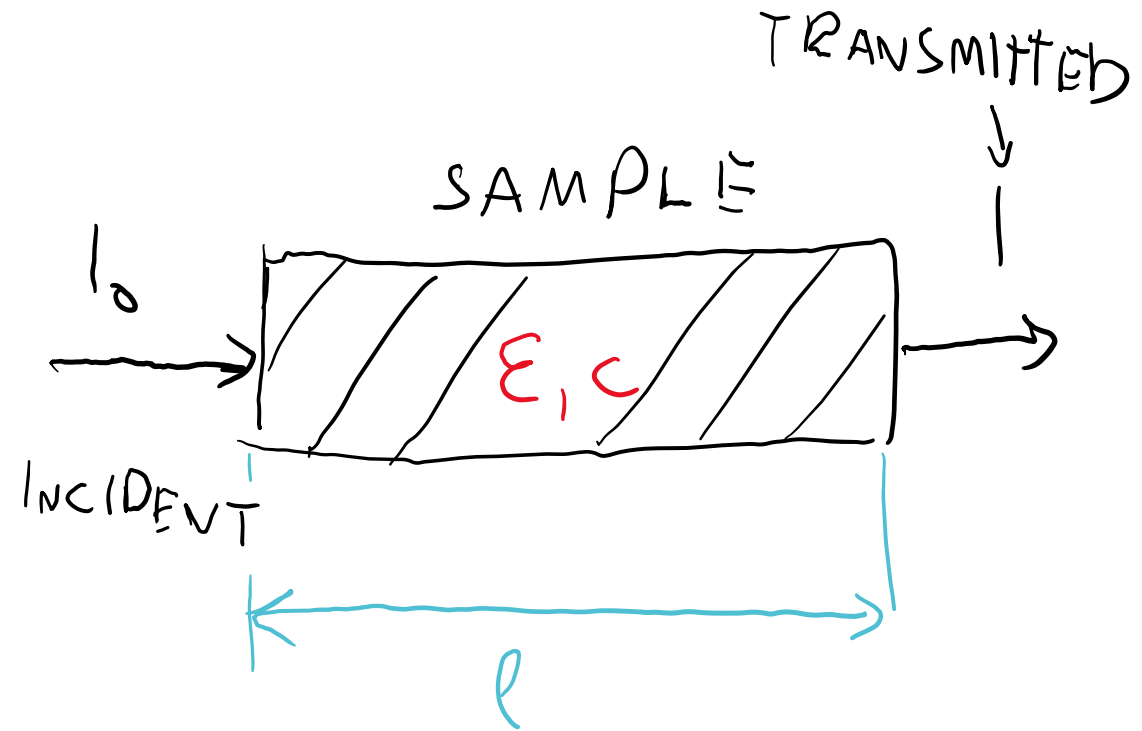
- Measurement modes:

- Absorbance

$$A = \frac{I_0}{I}$$

- Transmittance

$$T = \frac{I}{I_0}$$



$A$ : absorptivity [a.u.]

$\epsilon$ : molar attenuation coefficient  $\left[ \frac{\text{m}^2}{\text{mol}} \right]$

$l$ : optical path length [m]

$c$ : concentration of species  $\left[ \frac{\text{mol}}{\text{m}^3} \right]$



# SPECTROPHOTOMETRY

- Beer-Lambert Law:  
attenuation of light  $\Leftrightarrow$  properties of material being lit

$$A = \epsilon l c \text{ [a.u.]}$$

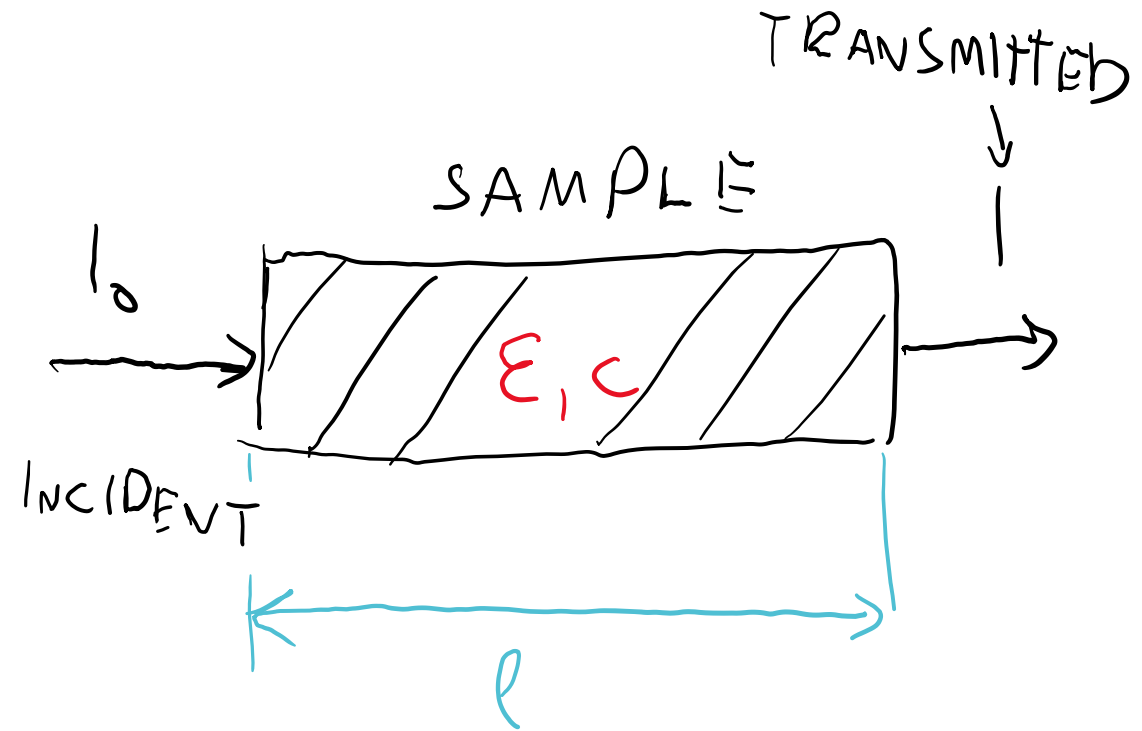
- Measurement modes:

- Absorbance

$$A = \frac{I_0}{I}$$

- Transmittance

$$T = \frac{I}{I_0}$$



$A$ : absorptivity [a.u.]

$\epsilon$ : molar attenuation coefficient  $\left[ \frac{\text{m}^2}{\text{mol}} \right]$

$l$ : optical path length [m]

$c$ : concentration of species  $\left[ \frac{\text{mol}}{\text{m}^3} \right]$





# SPECTROPHOTOMETRY

- Beer-Lambert Law:  
attenuation of light  $\Leftrightarrow$  properties of material being lit

$$A = \log_{10} \left( \frac{I_0}{I} \right) = \epsilon l c$$

- Measurement modes:

- **Absorbance**

$$A \approx \frac{I_0}{I}$$

- Transmittance

$$T = \frac{1}{A} \approx \frac{I}{I_0}$$

Transparent:

$$I = I_0$$

$$T \approx 100\%$$

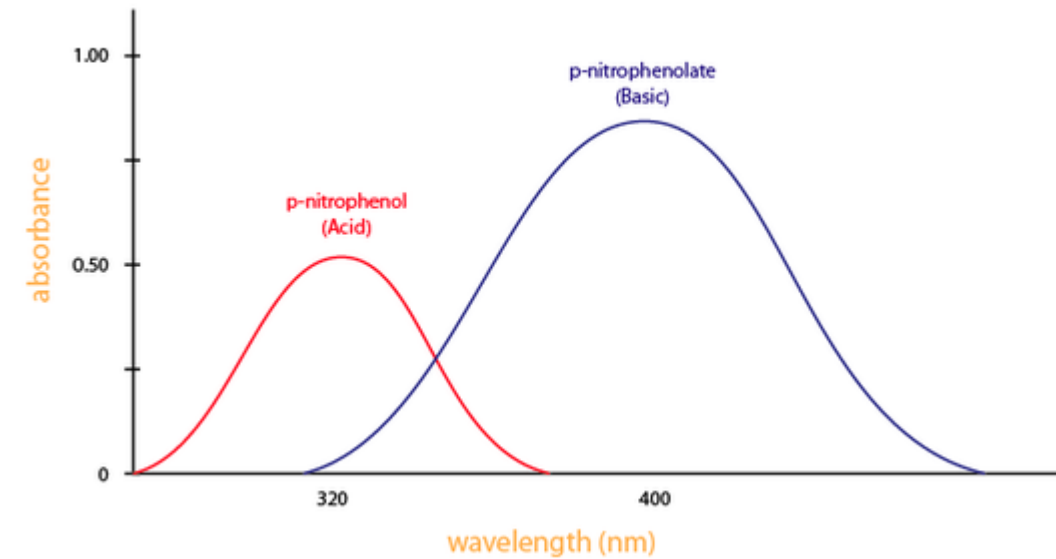
$$A \approx 0\%$$

Opaque:

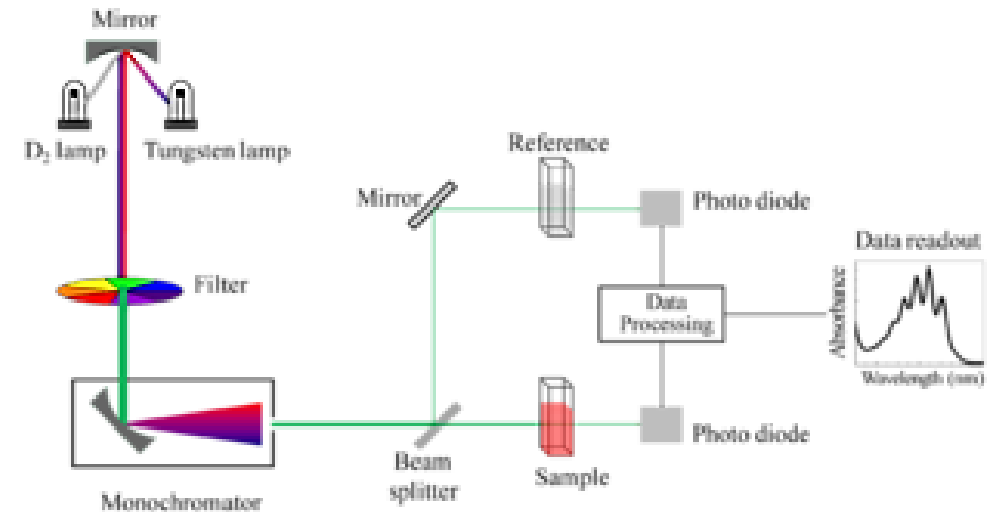
$$I \rightarrow 0$$

$$T \rightarrow 0\%$$

$$A \rightarrow 100\%$$



This Photo by Unknown Author is licensed under [CC BY-SA](#)



This Photo by Unknown Author is licensed under [CC BY-SA](#)



# FLUOROMETRY

- Fluorochrome/fluorophore: molecule that absorbs energy from incoming light and emits another light

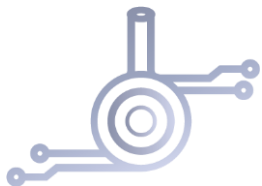
$$E = h \frac{c}{\lambda} [J]$$

- Where h: Planck's constant, c: speed of light [m/s],  $\lambda$ : light wavelength [m]

- Fluorescent intensity  $\Leftrightarrow$  lifetime

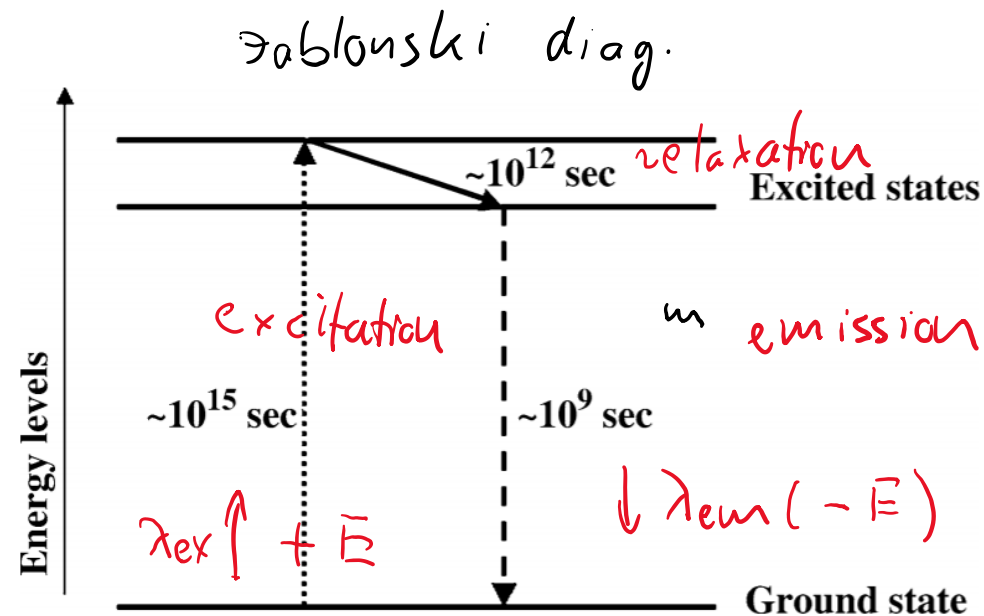
$$I(t) = I_0 e^{\left(-\frac{t}{\tau}\right)}$$

$\tau$ : fluorescent lifetime

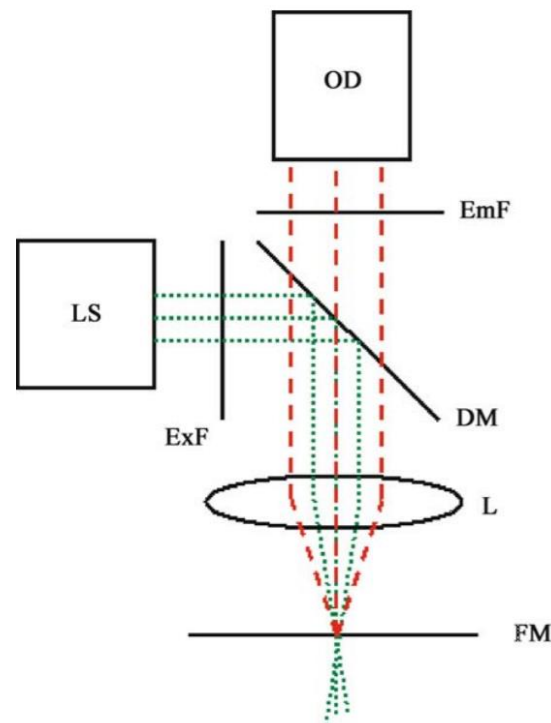


**TAL  
TECH**

TALLINN UNIVERSITY  
OF TECHNOLOGY



$$\lambda_{ex} < \lambda_{em}$$



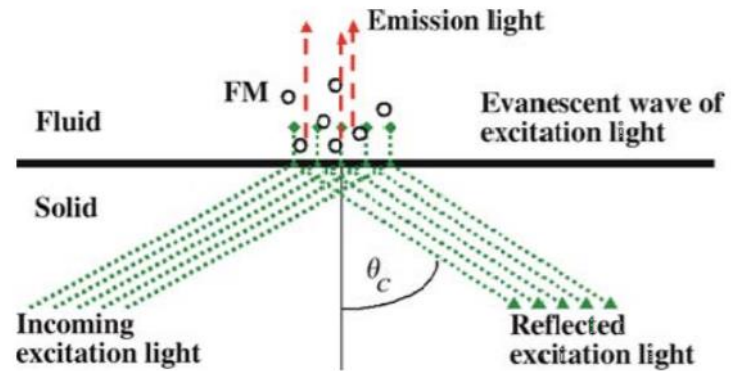
LS = light source,  
OD = optical detector,  
ExF = excitation filter,  
EmF = emission filter,  
DM = dichroic mirror,  
L = lens,  
FM = position of  
fluorochrome molecules



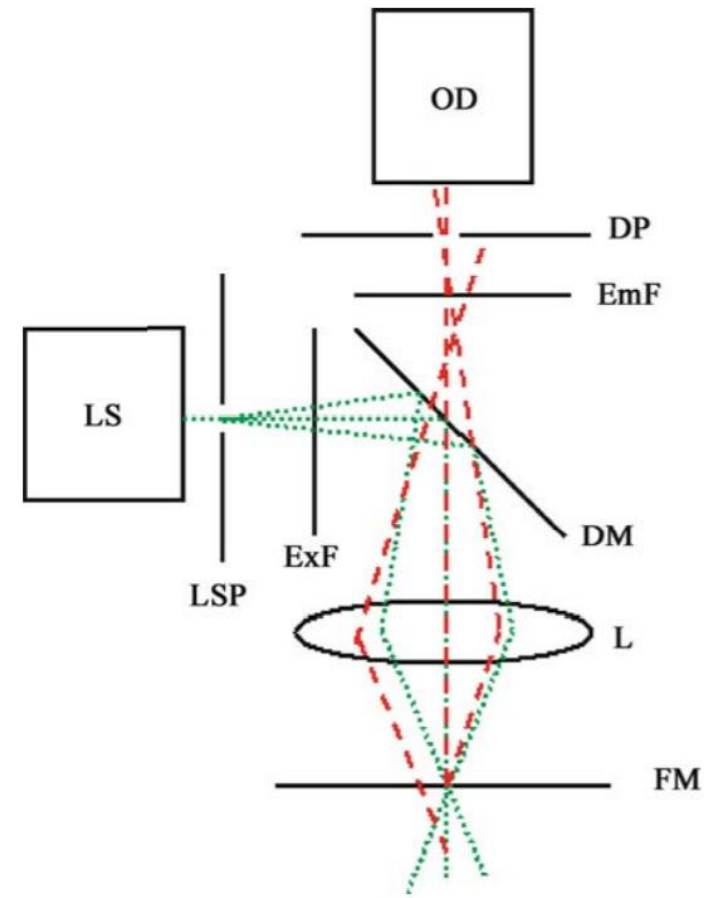
Optical

# FLUOROMETRY

- Special cases



**Fluorescence Measurements, Figure 4** The schematic concept of TIRF. FM = fluorochrome molecules. Note that only fluorochrome molecules that reside very close to the solid–fluid interface is illuminated by the excitation light and generates the emission light; the other fluorochrome molecules are not involved in the measurement



**Fluorescence Measurements, Figure 3** The schematic concept of a confocal fluorescence microscope configuration. LS = light source, OD = optical detector, ExF = excitation filter, EmF = emission filter, DM = dichroic mirror, L = lens, FM = position of fluorochrome molecules, LSP = Light-source pinhole, and DP = detector pinhole. Note that the emission light coming from out-of-focus fluorochrome molecules is blocked away by the detector pinhole

# AMPEROMETRY

- Faraday's first law of electrolysis:  
"the amount of chemical change produced by current at an electrode-electrolyte boundary is proportional to the quantity of electricity used"
- Reaction: reduction-oxidation (redox)
- Electrodes:**
  - Working** (where the reaction e.g. oxidation happens, material e.g. C, Au)
  - Auxiliary** (counter, where the reaction goes in the opposite direction, e.g. reduction, material e.g. Pt)
  - Reference** (Ag/AgCl)
- Amperometry:**
  - Voltage applied to working electrode
  - Current measured from working to counter electrode

Faraday's law of electrolysis:

$$i_t = \frac{\partial Q}{\partial t} = \eta F \frac{\partial N}{\partial t}$$

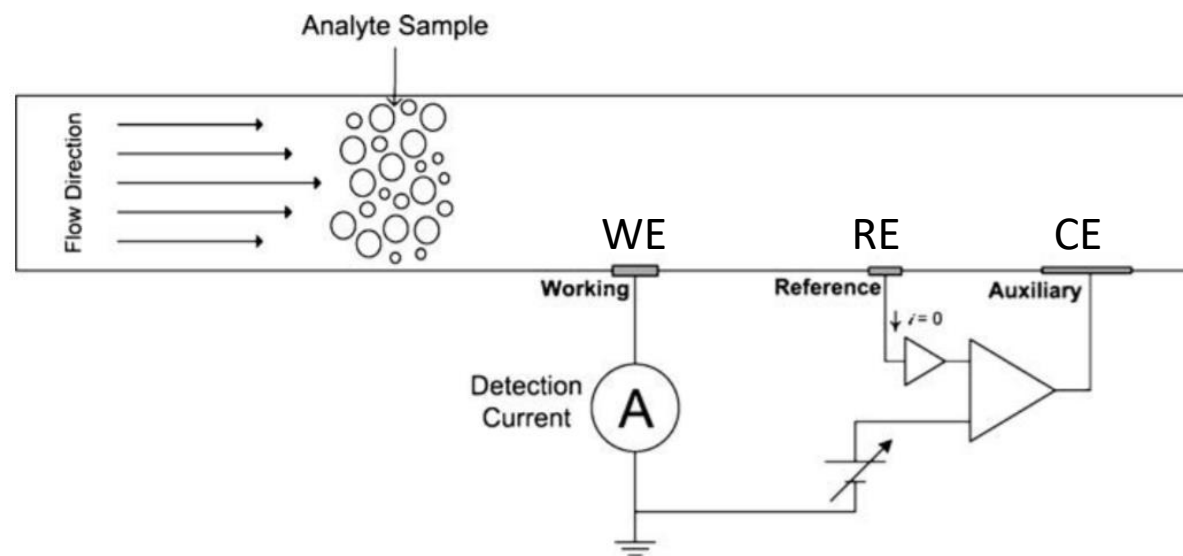
$i_t$ : current generated at time  $t$  [A]

$Q$ : charge [C]

$\eta$ : valency

$N$ : moles of analyte [mol]

$F$ : Faraday constant



# AMPEROMETRY

- Faraday's first law of electrolysis:  
"the amount of chemical change produced by current at an electrode-electrolyte boundary is proportional to the quantity of electricity used"
- Reaction: reduction-oxidation (redox)
- **Electrodes:**
  - **Working** (where the reaction e.g. oxidation happens, material e.g. C, Au)
  - **Auxiliary** (counter, where the reaction goes in the opposite direction, e.g. reduction, material e.g. Pt)
  - **Reference** (Ag/AgCl)
- **Amperometry:**
  - Voltage applied to working electrode
  - Current measured from working to counter electrode

Faraday's law of electrolysis:

$$i_t = \frac{\partial Q}{\partial t} = \eta F \frac{\partial N}{\partial t}$$

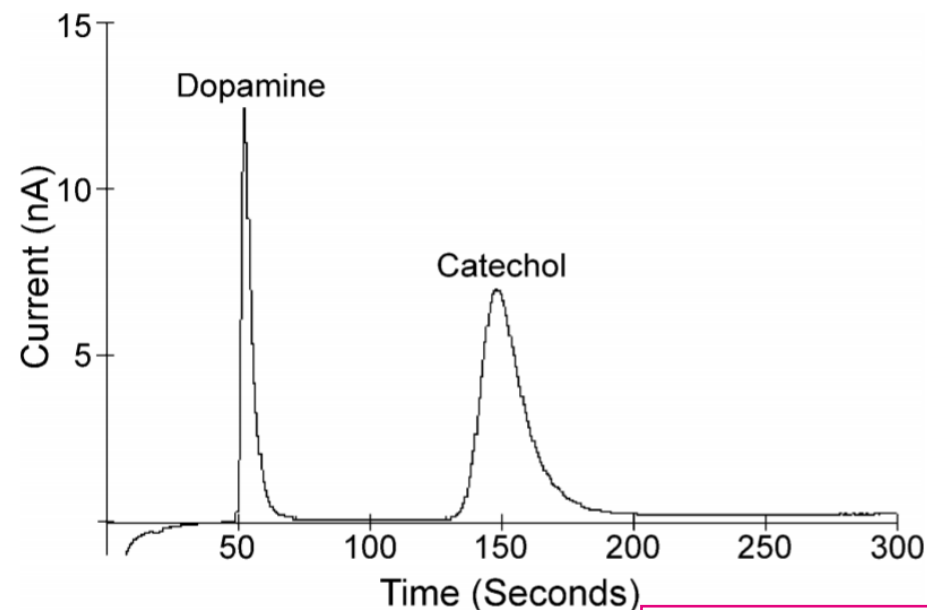
$i_t$ : current generated at time  $t$  [A]

$Q$ : charge [C]

$\eta$ : valency

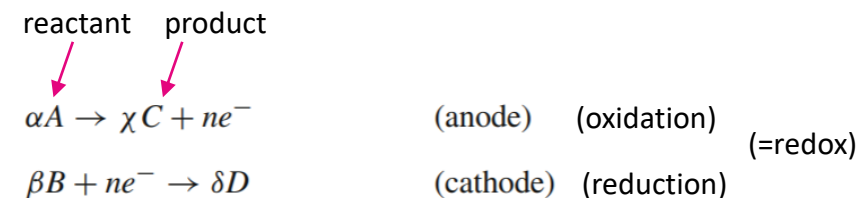
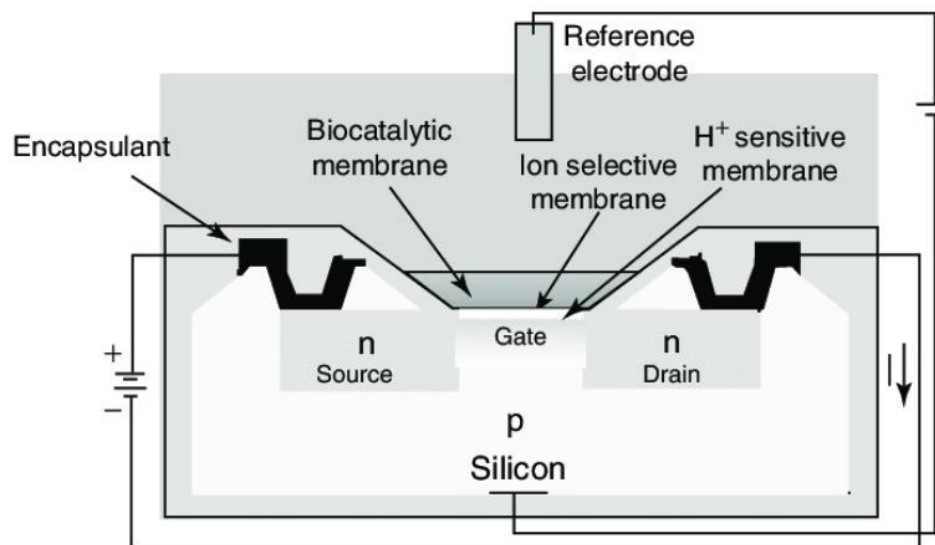
$N$ : moles of analyte [mol]

$F$ : Faraday constant



# POTENTIOMETRY

- Setup similar to amperometry, but
  - Electrode impedance high → minimal current flow
  - Charge accumulation on electrode → Nernst potential measured



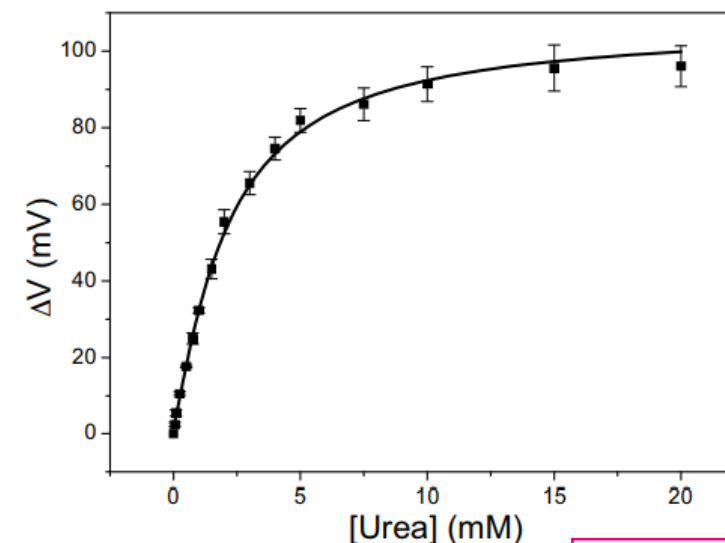
$$E = E^0 - \frac{0.059}{|n|} \log \left( \frac{a_C^\chi a_D^\delta}{a_A^\alpha a_B^\beta} \right)$$

$E$ : working electrode potential [V]

$E^0$ : standard electrode potential

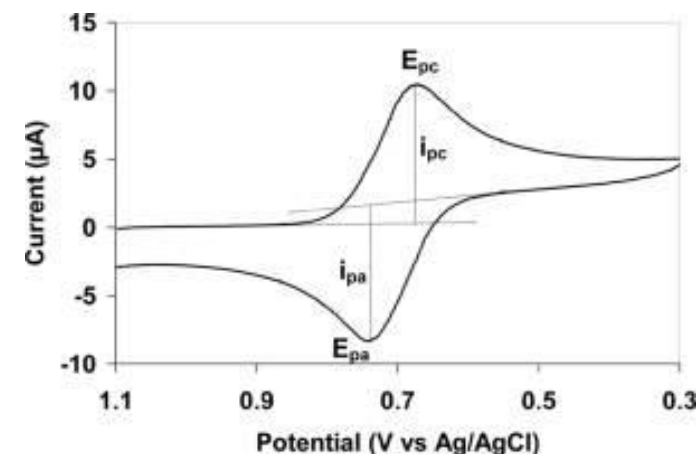
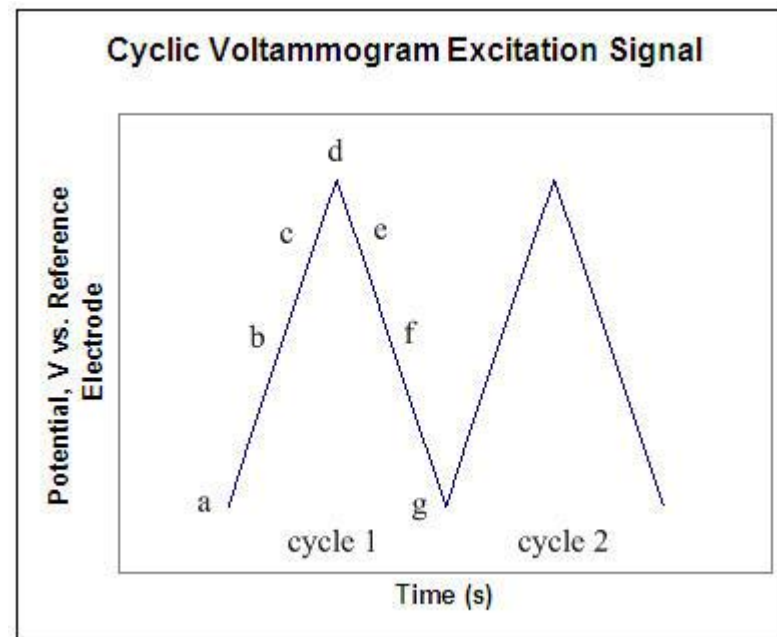
$n$ : number of  $e^-$

$a$ : activity of A – D



# CYCLIC VOLTAMMETRY

- Working electrode potential linearly and cyclically ramped in time
- Electrode potential:  $E = E_i + vt$ 
  - $E_i$ : initial potential [V]
  - $v$ : sweep rate  $\left[\frac{V}{s}\right]$
  - $t$ : time [s]
- Concentration (Fick's 2<sup>nd</sup> law):
  - $\frac{\partial c}{\partial t} = D \frac{\partial^2 c}{\partial x^2}$
  - Concentration change <-> diffusion** (to electrode surface)

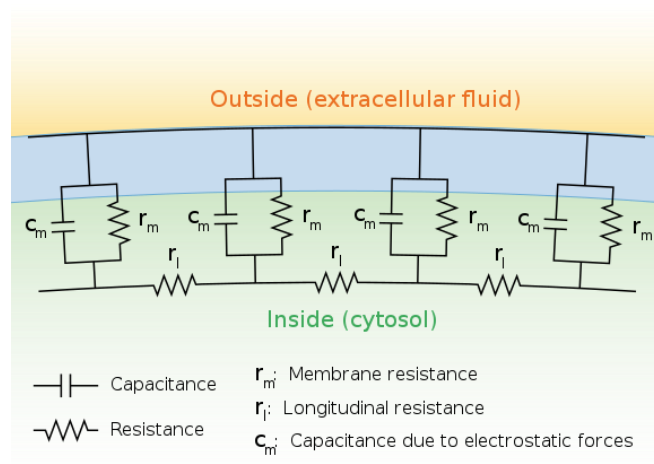


This Photo by Unknown Author is licensed under [CC BY-SA](https://creativecommons.org/licenses/by-sa/4.0/)



# IMPEDANCE SPECTROSCOPY

- Tissue  $\sim$  resistor(s) + capacitor(s)
- If we use A.C. excitation, e.g. sinusoidal, complex impedance will change according to the excitation frequency
- Cells:



[This Photo](#) by Unknown Author is licensed under [CC BY-SA](#)

*Resistor:*

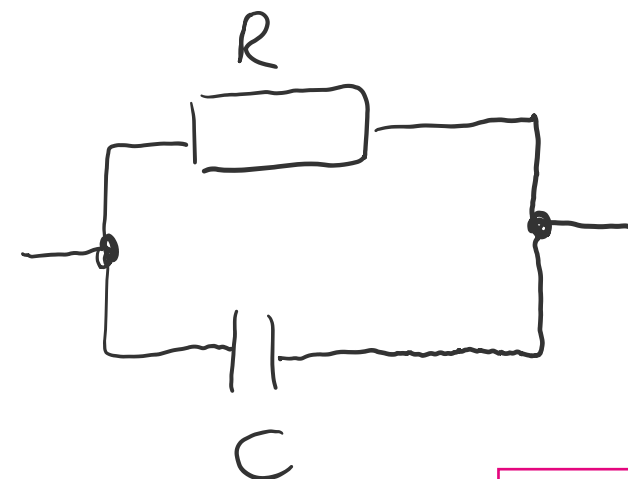
$$Z_R = \frac{V(t)}{I(t)} = \text{Re}(Z) - j \cdot \text{Im}(Z)$$

*Capacitor:*

$$Z_C = \frac{1}{j\omega C}$$

$\omega$ : angular freq.

$\omega = 2\pi f$  where  $f$ : excitation freq.



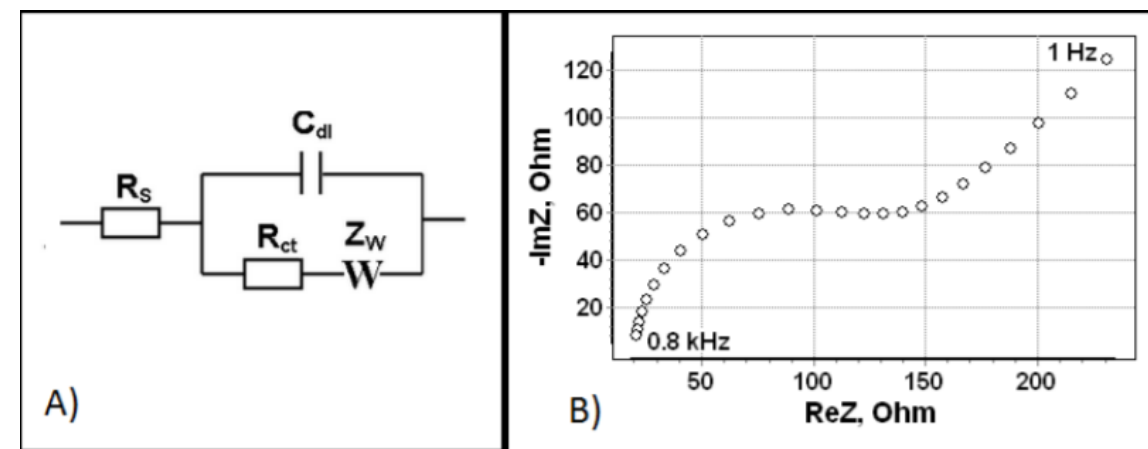
# IMPEDANCE SPECTROSCOPY

- Tissue  $\sim$  impedance(s) + capacitance(s)
- We measure varying impedance in response to voltage change
- Setup:
  - working + counter electrode + max. 2 reference electrodes
  - Randles equivalent circuit
  - Bode or Nyquist plot used for visualization
  - At high frequency:  $I = V/R_S$
  - At mid to low frequency:  $I = V/(R_S + R_{CT})$

$$Z^* = \frac{V(t)}{I(t)} = \text{Re}(Z) - j \cdot \text{Im}(Z)$$

$$I(t) = V(t) \frac{1 + (\omega RC)^2}{R - j\omega R^2 C}$$

$\omega = 2\pi f$  where  $f$ : excitation freq.



$R_S$ : electrolyte resistance  
 $C_{dl}$ : double – layer capacitance  
 $R_{ct}$ : charge – transfer resistance  
 $Z_W$ : Warburg element

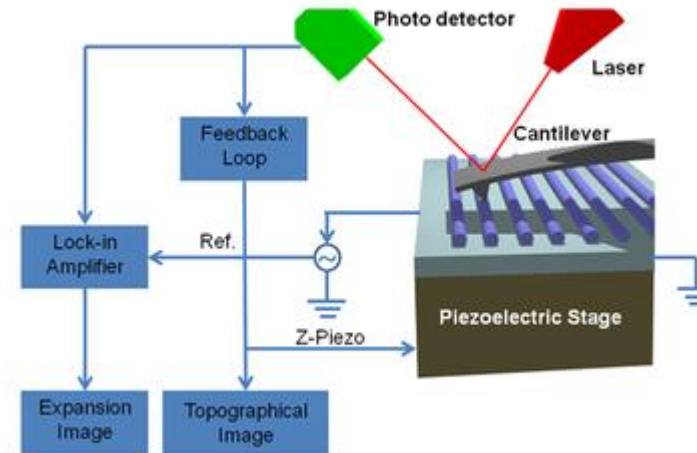
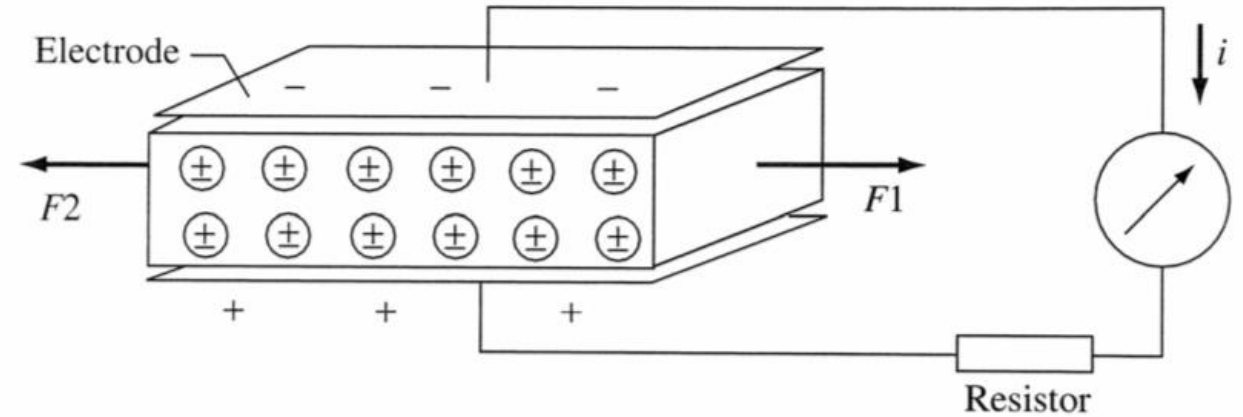
$R_S = 20 \, \Omega$ ,  $C_{dl} = 25 \, \mu\text{F}$ ,  $R_{ct} = 100 \, \Omega$ ,  $A_W = 300 \, \Omega \cdot \text{s}^{-0.5}$

# PIEZOELECTRIC SENSORS

- Electromechanical sensors
- Piezoelectric effect:
  - deformation  $\Leftrightarrow$  electric field

$$S = dE$$
$$D = dT$$

- Where:
  - $S$ : strain
  - $d$ : piezoelectric coeff.
  - $E$ : electric field  $\left[\frac{V}{m}\right]$
  - $D$ : linear displacement  $[m]$
  - $T$ : stress  $[N/m^2]$



This Photo by Unknown Author is licensed under [CC BY-SA](#)

This Photo by Unknown Author is licensed under [CC BY-SA](#)

# CANTILEVERS

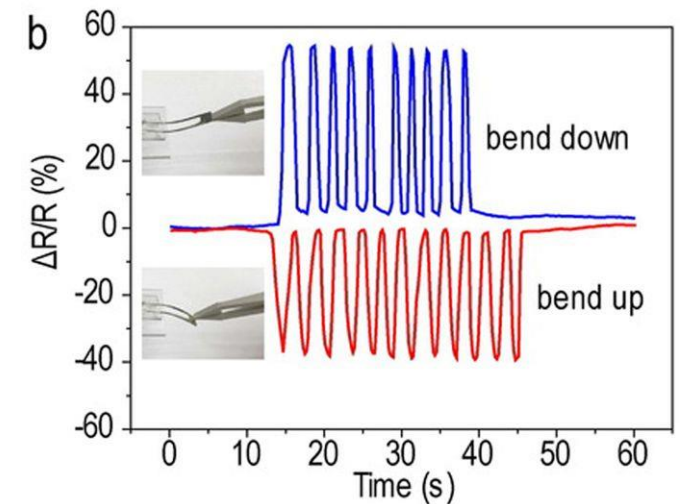
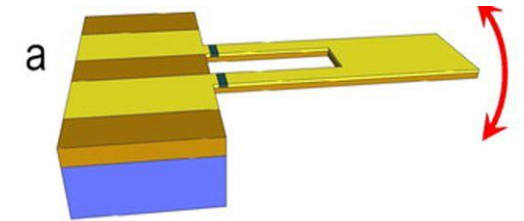
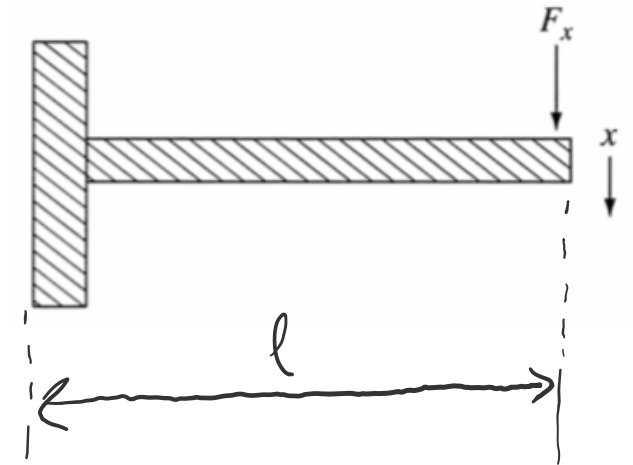
- Typically microcantilevers
- Beam displacement  $x \leftrightarrow$  Applied force  $F_x$  and beam length  $l$

$$\Delta x = \frac{l^3}{3E_m I_m} F_x$$

$$F_x = k_m \Delta x$$

- Where:

- $E_m$ : Young's modulus  $\left[\frac{N}{m^2}\right]$
- $I_m$ : second moment of inertia  $[kg/m^2]$
- $F_x$ : Force  $[N]$
- $l$ : length  $[m]$
- $k_m$ : spring constant  $\left[\frac{N}{m}\right]$



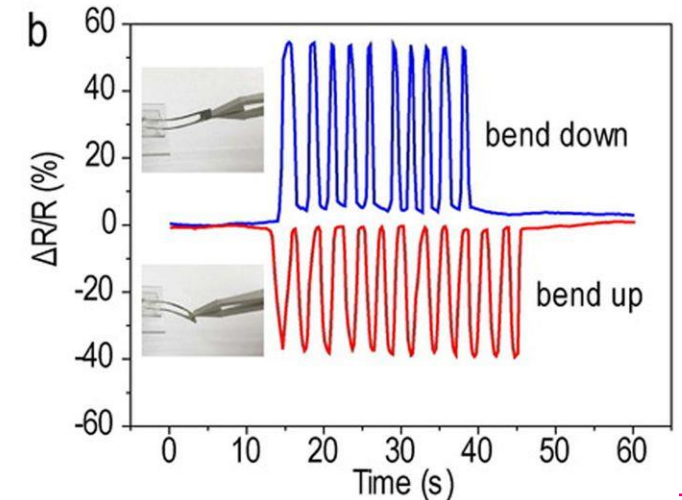
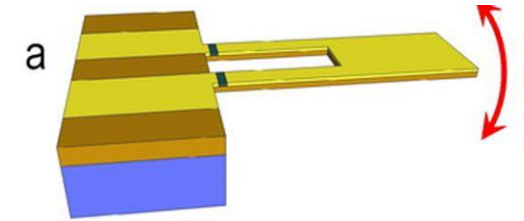
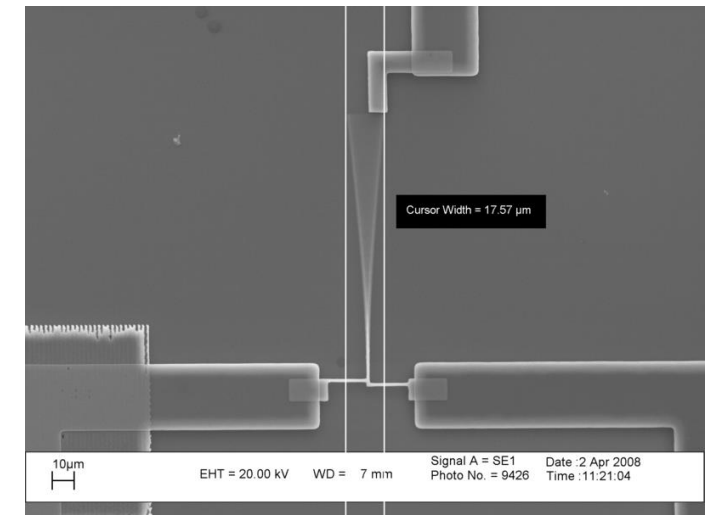
# CANTILEVERS

- Typically microcantilevers
- Beam displacement  $x \leftrightarrow$  Applied force  $F_x$  and beam length  $l$

$$\Delta x = \frac{l^3}{3E_m I_m} F_x$$

$$F_x = k_m \Delta x$$

- Means to convert mechanical to electrical:
  - Strain gauge (resistance  $\leftrightarrow$  force)
  - Optical (diffraction  $\leftrightarrow$  force)
  - Piezoelectric

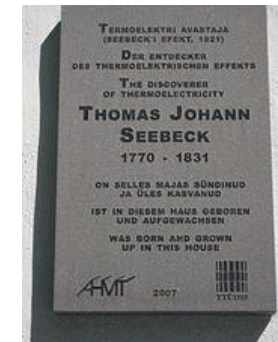
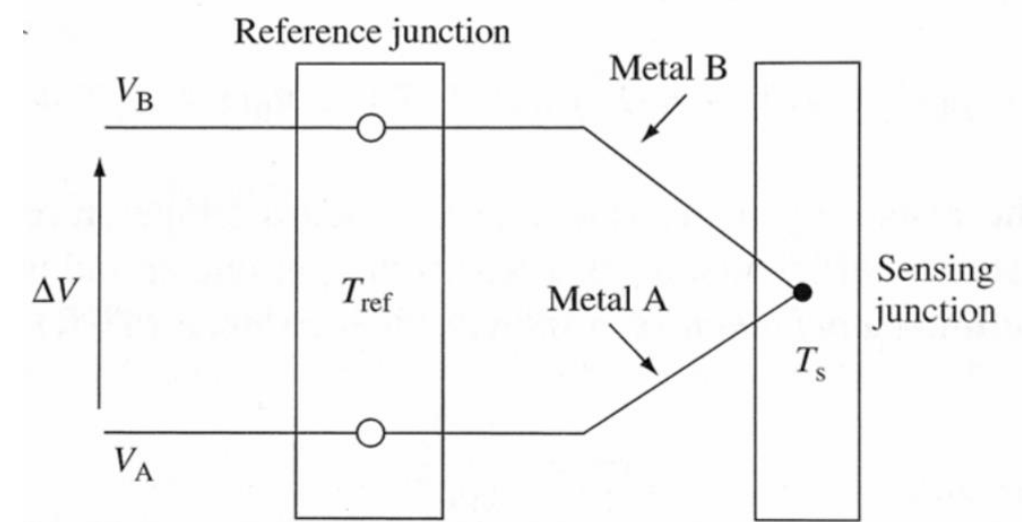


# THERMOCOUPLE

- Based on the Seebeck effect:
- Temperature difference at a junction of dissimilar conductors creates an electric potential difference:

$$\Delta V = \alpha_S \Delta T$$

- $\Delta V$ : electrical voltage [V]
- $\alpha_S$ : Seebeck coefficient  $\left[\frac{V}{K}\right]$
- $\Delta T$ : temperature difference



## WHAT WE'VE COVERED IN THIS LECTURE

Detection method	Example	Measured response to analyte concentration	Pros	Cons
Optical	Spectrophotometry	Optical intensity change	No electrical interference, contactless	Slow response, complex instrumentation
Electrical	Cyclic voltammetry	Current change in response to voltage input	Highly sensitive and compact, simple fabrication	Sensitive to noise
Mechanical	Cantilevers	Beam deflection (analyte mass is detected)	Highly sensitive and compact	Complex fabrication, sensitive to vibrations





# SENSORS

- Different types of sensors: EM, Mechanical, Chemical
- Detection mechanisms
- Working principles

01.02.2022



# TAL TECH

## IEE1860 BIOMEMS

Contact: [tamas.pardy@taltech.ee](mailto:tamas.pardy@taltech.ee)

**TALLINN UNIVERSITY  
OF TECHNOLOGY**



**TAL  
TECH**

# BIOMEMS

**self**  
**diagnostics**

